

Extracellular vesicle-mediated transfer of constitutively active MyD88^{L265P} engages MyD88^{wt} and activates signaling

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Running Title: EV-transmitted MyD88^{L265P} promotes inflammation

Key points

- MyD88^{L265P} is present in the extracellular vesicles (EVs) secreted by WM cancer cells and triggers signaling in the recipient cells
- MyD88-containing EVs shape proinflammatory microenvironment in the bone marrow

Abstract

The link between inflammation and cancer is particularly strong in Waldenström's macroglobulinaemia (WM), a diffuse large B-cell lymphoma, wherein the majority of patients harbor a constitutively active mutation in the innate immune signaling adaptor MyD88. MyD88^{L265P} constitutively triggers the myddosome assembly providing a survival signal for cancer cells. Here we report detection and a functional role of MyD88 in the extracellular vesicles (EVs) shed from WM cells. MyD88^{L265P} was transferred via EVs into the cytoplasm of the recipient mast cells and macrophages, recruiting the endogenous MyD88 that triggered the activation of proinflammatory signaling in the absence of receptor activation. Additionally, internalization of EVs containing MyD88^{L265P} was observed in mice with an effect on the bone marrow microenvironment. MyD88-loaded EVs were detected in the bone marrow aspirates of WM patients thus establishing the physiological role of EVs for MyD88^{L265P} transmission and shaping of the proinflammatory microenvironment. Results establish the mechanism of transmission of signaling complexes via EVs to propagate inflammation as a new mechanism of intercellular communication.

Introduction

MyD88 is a signaling adapter protein that plays a pivotal role in innate immunity. MyD88 is recruited to the TIR domains of activated Toll-like receptors (TLRs) and interleukin-1/18 receptor (IL-1R, IL-18R), leading to myddosome complex formation with IRAK4 and IRAK1/2 kinases.^{1,2} Myddosome activates pathways resulting in activation of transcription factors NF- κ B, AP-1, IRFs and others, which induce the transcription of numerous proinflammatory genes.³ Patients with germline *MYD88* loss-of-function mutations only survive into adulthood on strict therapy of antibiotics.⁴ Conversely, somatic *MYD88* gain-of-function mutations within the TIR domain of MyD88 enable constitutive activation.⁵⁻⁷ MyD88Leu265Pro (MyD88^{L265P}) mutation triggers spontaneous assembly of the myddosomal complex with IRAK kinases resulting in activation of STAT3, NF- κ B, secretion of IL-6, IL-10 and IFN β .^{5,8} Activating mutations in *MYD88* are frequently detected in ABC subtype of DLBCL (diffused large B cell lymphoma), chronic lymphocytic leukemia and are present in more than 90% of patients with WM (Waldenström`s macroglobulinaemia).^{9,10} MyD88^{L265P} supports the survival of lymphoma cells by upregulating Bcl-xL expression through NF- κ B pathway¹¹ and activation of Bruton`s tyrosine kinase (BTK).¹² Inhibition of the signaling pathway downstream of MyD88 reduces proliferation of cancer cell.⁵⁻⁷

The role of extracellular vesicles (EVs; comprising microvesicles (MVs) and exosomes) has been described in cancer development and dissemination of signals to other cells.¹³ The number of EVs increases in stress conditions (reviewed in^{14,15}) and increased EV levels were detected in the peripheral blood of cancer patients.¹⁶ EVs encapsulate different cellular components including proteins, lipids, carbohydrates and noncoding regulatory RNAs, such as miRNA. Tumor EVs promote directional cell motility through extracellular matrix (ECM)¹⁷ and ECM remodeling.^{18,19} They also alter physiology of both surrounding and distant nontumor cells, which in turn allows growth and dissemination of cancer cells.^{20,21} However, transmission of

the proteins of signaling pathways which could directly activate the intracellular signaling pathway of recipient cells has not been described.

EVs are enriched in membrane-associated proteins that form aggregates.²² Mutant MyD88^{L265P} has an intrinsic aggregation propensity and is found associated to the cellular membranes, thus it fulfils both of the enrichment criteria. We therefore surmised that MyD88 might be associated to the EVs released by lymphoma cells. This may be relevant if the transferred MyD88 mutant could trigger cell activation in recipient cells that internalize the EVs. Previous proteomic analysis of the exosomes from human thymic tissue identified MyD88 among many other protein components²³, giving credence to the hypothesis.

While WM cells can influence their microenvironment through direct interaction with stromal cells, and by the release of cytokines^{24,25} in this report a mechanism of intercellular proinflammatory signaling mediated by EVs was investigated. Here we show that the EVs from the lymphoma cell lines as well EVs isolated from the bone marrow aspirates of WM patients contained MyD88. MyD88^{L265P}-containing EVs are internalized by the recipient cells, where the transferred MyD88^{L265P} recruits the MyD88^{wt} and triggers activation of the inflammatory pathway thus propagating the inflammatory signaling independent of the TLR and IL-1R receptors. EVs with MyD88^{L265P} injected i.v. were able to activate the reporter cells in mice thus demonstrating the possibility of the *in vivo* transfer of the functional signaling complexes. EVs that were injected into the bone marrow increased globulin concentration in blood and modified the bone marrow microenvironment by recruitment/differentiation of mast cells, which support the development and propagation of B cell lymphomas. These results extend our understanding how cancer cells can modify the tumor's microenvironment and identified a mechanism of the transmission of signaling components via EVs as an additional mechanism of intercellular communication.

Methods

Materials

LY294002 (Invivogen), Dynasore (Sigma), LPS (from *Salmonella abortus equi* HL83 - provided by K. Brandenburg; Forschungszentrum Borstel, Germany); Antibodies against: MyD88, IRAK4 (Cell Signaling Technology), Tsg101 (Santa Cruz Biotechnology), GAPDH and β -actin (Cell Signaling Technology), secondary HRP-conjugated anti-mouse (Santa Cruz Biotechnology) or anti-rabbit antibody (Abcam).

Cell lines

HEK293 cell line (human embryonic kidney) from ATTC, HEK293 MyD88^{KO} and HEK293 NEMO^{KO} cell lines prepared with CRISPR/Cas9 (a gift from D. Abbott), mouse immortalized bone-marrow derived macrophages (iBMDMs) (a gift from K. Fitzgerald); MWCL-1²⁶ and BCWM.1²⁷ cell lines established from the bone marrow aspirate of a patient with WM and Ramos-Blue cell line (Invivogen) were used in these studies.

Mice

C57BL/6 (wt and IRAK4^{KO}), BALB/c and SCID (C.B-17/lctHsd-*Prkdc*^{scid}*Lyst*^{bg-J}) mice were purchased from Harlan (Italy). 8-12 weeks old male and female mice were used for experiments. No sample size estimation was performed. All the animal samples were included in the analysis. No blinding or randomization was performed. All animal experiments were performed according to the directives of the EU 2010/63 and were approved by the Administration of the Republic of Slovenia for Food Safety, Veterinary and Plant Protection

of the Ministry of Agriculture, Forestry and Foods, Republic of Slovenia (no. U34401-3/2017/8).

Bone marrow derived-cultured mast cells (BMCMCs) were prepared from bone marrow cells from C57BL/6 mice using 10 ng/ml of recombinant mIL-3 (PeproTech) for 4 weeks. BMDMs were prepared from bone marrow cells from C57BL/6 mice using 40 ng/ml M-CSF for 4 days.

Preparation of EVs

HEK293 cells were seeded in DMEM + 10% exosome depleted FBS (System Biosciences) and transfected with MyD88^{L265P}. After 24 h supernatants were collected. MWCL-1, BCWM.1 or Ramos cells were seeded at density 2×10^7 cells in 20 ml of RPMI + 10% exosome depleted FBS. After 48 h supernatants were collected. To prepare EV_{CONT} supernatants from growing HEK293 cells were collected. Supernatants were centrifuged at 800 g for 15 min to remove the cells and at 100 000 g for 1h to isolate the EVs. EVs were washed with PBS and ultracentrifuged again. EVs were resuspended in PBS and protein concentration was measured using BCA assay (Sigma).

Use of patient bone marrow aspirates was obtained after informed consent approved by the institutional review board at Dana-Farber Cancer Institute. This study was conducted in accordance with the Declaration of Helsinki. Bone marrow aspirates from patients with WM were centrifuged and serum was separated. 1 drop of EDTA/5ml was added, gently mixed and left at 4°C overnight. To remove any precipitation samples were centrifuged at 300 g for 10 min then again at 2000 g for 10 min. To pellet the MVs supernatants were centrifuged at 16 000 g for 20 min. Supernatants were filtered through 0.2 µm filters and ultracentrifuged at 120

000 g for 1 h to isolate exosomes. The pellet was washed with PBS and ultracentrifuged again. Exosomes and MVs were resuspended in PBS and stored at -80° C.

NF- κ B luciferase reporter assay

HEK293 or HEK293 MyD88^{KO} cells were transfected with firefly luciferase under NF- κ B promotor and *Renilla* luciferase reporter plasmids. Cells were stimulated with EVs and luminescence was measured on Orion luminometer (Berthold).

Quantitative PCR

BMCMCs, iBMDMs or BMDMs were stimulated with EVs. RNA was isolated using Trizol (Roche) or Purelink RNA mini kit (Invitrogen), cDNA was prepared using High capacity cDNA reverse transcription kit (Applied Biosystems) and qPCRs for mouse *Gapdh*, *Rantes* or *Il6* were performed using SYBR green I master kit (Roche) on LightCycler 480 (Roche).

Immuno-electron microscopy

EVs were fixed with 4% formaldehyde in 0.1 M phosphate buffer and pelleted by centrifugation at 200 g. The pellet was washed with PBS/0.15% glycine, embedded in 12% gelatin and cut into 0.5×0.5×0.5 mm blocks. The blocks were cryoprotected with 2.3 M sucrose, mounted on specimen holders and frozen in liquid nitrogen. Ultrathin cryo-sections (50 nm thick) were cut at -120°C. Sections were retrieved with 1:1 mixture of 2.3 M sucrose and 2% methyl cellulose on Au grids. Unspecific labeling was blocked with 1% BSA-cTM (Aurion). Sections were incubated with anti-Myd88 antibody and then with protein A

conjugated to 10 nm gold (UMC Utrecht). Sections were embedded and stained with methyl cellulose/uranyl acetate and examined in a Philips CM100 transmission electron microscope at 80 kV.

Confocal microscopy

HEK293 or HEK293 MyD88^{KO} cells were left untreated or transfected with MyD88^{wt-YFP}. EV_{L265P-CFP} or EV_{L265P-YFP} were added to the cells. Cell membranes were dyed with cholera toxin B (CT-B)-Alexa647 (Invitrogen). Cells were observed using Leica TSC SP5 confocal microscope equipped with an HCX plan apo 963 (NA 1.4) oil immersion objective. For acquisition and image processing, Leica LAS AF software was used.

In vivo internalization of EVs

HEK293 cells were transfected with firefly luciferase under NF- κ B promotor. After 24 h cells were collected. 2×10^6 cells/mouse were s.c. injected into right flank of BALB/c mice. Afterwards 150 μ g/mouse of EV_{L265P} or EV_{CONT} were injected i.v. After 24 h mice were anesthetized with isoflurane and D-luciferin (Xenogen) 150 mg/kg of body weight was injected s.c. into left flank. After 10-15 min mice were imaged *in vivo* using IVIS Lumina Series III (Perkin Elmer). Data were analyzed with Living Image 4.5.2 (Perkin Elmer). Three mice were used in two separate experiments for each measurement.

Injection of EVs into the bone marrow and analysis

PKH67-labelled EVs were intramedullary injected into femur of C57BL/6 mice as described in Zilber et al.²⁸ After 16 h bones were collected and PKH fluorescence in cells was detected using CyFlow space flow cytometer (Partec). EV_{MWCL}, EV_{CONT} or PBS were intramedullary injected. After 8 days mice were sacrificed, blood was collected and serum was prepared (Sarstedt; 3000 rpm/30 min). Blood profile was determined from sera using VetScan comprehensive diagnostic profile reagent rotor and analyzed on VetScan VS2 (Abaxis). For rouleaux formation blood smears were prepared from a drop of whole blood using Hemacolor rapid staining of blood smear kit (Merck). Femurs were collected for immunohistochemistry. Bones were decalcinated using 9% formic acid and embedded in paraffin (Leica Paraplast). The paraffin blocks were cut 7 µm thick with a rotation microtome RM 2245 (Leica). Mast cells were labelled using antiCD40L (CD154) (Biolegend) and detected by HRP-conjugated anti-goat armenian hamster (Abcam) using TMB reagent (Sigma) under Leica DMi8 with Leica MC170 HD camera.

Statistical analysis

Representative experiments are shown. For the analysis of experimental data, a *t*-test was used. **P* < 0.1; ***P* < 0.05; ****P* < 0.01; *****P* < 0.005; n.s. – not significant.

Results

Presence of the MyD88 in EVs and transmission of the signal to the recipient cells

For the initial characterization of the ability of the MyD88^{L265P} mutant to be incorporated into the vesicles, the EVs were isolated from the supernatant of unstimulated MyD88^{L265P}

transfected HEK293 cells and characterized by the dynamic light scattering (DLS) (supplemental Figure 1A), which showed the presence of MVs and exosomes. As shown by WB, EV_{L265P} contained MyD88, which was absent from the EV_{CONT}, isolated from the control HEK293 cells where both types of EVs contained exosomal marker Tsg101 (Figure 1A). In order to show that the EVs carrying the constitutively active MyD88^{L265P} can be taken up by the recipient cells and that this form of the delivery of MyD88 is sufficient to trigger activation of the signaling pathway in the recipient cells, activation of the NF-κB-dependent luciferase reporter was monitored in HEK293 cells. Addition of EV_{L265P} but not EV_{CONT} to the reporter cells activated NF-κB (Figure 1C) thus supporting the hypothesis that the constitutively active signaling protein can be transmitted to the recipient cells, where it can activate the signaling pathway.

MWCL-1 is a WM cell line established from the bone marrow aspirate of a WM patient.²⁶ We observed that MWCL-1 cells which harbor the constitutively active MyD88^{L265P} mutation release high amount of EVs. EV_{MWCL1} were isolated from the supernatant and compared to the EV_{L265P}. As shown by DLS and TEM MWCL-1 cell line secreted EVs (exosomes and MVs) of similar size to those from HEK293 cell line (supplemental Figure 1B-C). WB revealed the presence of MyD88 in EV_{MWCL1}, along with the exosomal marker Tsg101 (Figure 1D) and was not present in the cell supernatant after ultracentrifugation (supplemental Figure 1D). Moreover, the presence of MyD88 was confirmed inside MVs by the immuno-electron microscopy (Figure 1E). Importantly, IRAK4, which is a constituent of the active myddosomal complex, was also detected in the EV_{MWCL1} (Figure 1F) though it was hardly detected in EV_{L265P} (Figure 1B). Furthermore, EV_{MWCL1} triggered activation of the NF-κB mediated signaling in the recipient cells (Figure 1G), demonstrating that cancer cells can indeed release EVs with inflammatory properties.

Internalization of the intact EVs is necessary for the activation of signaling in the recipient cells

Cytokines can be released via EVs thus inducing stimulation of cells. MWCL-1, but not Ramos cells (containing MyD88^{wt}), secreted IL-6, IL-8, IL-10, IL-12 and TNF α into the supernatant (supplemental Figure 2A), of which only IL-6 and IL-8 were detected in the EV_{MWCL1} (supplemental Figure 2B). IL-1 β , which signals via the MyD88 and could eventually stimulate the NF- κ B in a positive feedback loop, was not detected. MyD88^{L265P} constitutive activity promotes NF- κ B dependent cytokine expression. In order to rule out cytokine-triggered cell activation by EVs, the EVs were isolated from NEMO^{KO} (IKK γ) HEK293 cells²⁹ (supplemental Figure 3A-B) transfected with MyD88^{L265P}, where the NF- κ B signaling pathway and production of cytokines is inactivated. EV_{L265P_NEMOwt} and EV_{L265P_NEMO-KO} activated cells to a similar extent (Figure 2A), confirming that the activity is due to the EV-transmitted MyD88. Additionally, IRAK4^{KO} BMDMs were stimulated with EV_{L265P}, since MyD88 signaling pathway requires IRAK4 signaling kinase. Results demonstrate that *Rantes* mRNA expression almost completely diminished in IRAK4-deficient cells (Figure 2B), further confirming that MyD88 signaling pathway is indeed involved in the activation by the EVs.

EV_{L265P} were submitted to several freeze/thaw cycles to disrupt the membranes of the EVs. Freezing substantially decreased the activity (Figure 2C) demonstrating that the intact EVs and their content are necessary for the transfer of the signaling activity. Several mechanisms of EV-mediated transfer of molecular cargo have been described.³⁰⁻³² Dynasore, an inhibitor of dynamin-mediated endocytosis, partially inhibited NF- κ B activation triggered by EV_{L265P} (Figure 2D). Additionally, partial inhibition of the reporter cell activation was also observed after the addition of LY294002, a PI3K inhibitor (Figure 2E), which inhibits phagocytosis.

Results therefore suggest likely involvement of several mechanisms that contribute to the EV mediated signaling cargo internalization.

LPS induced signaling activates BTK, which interacts with several TIR domain containing proteins, including MyD88.³³ MyD88^{L265P} supports activation of BTK¹² in WM and secretion of the proinflammatory cytokines.²⁵ BTK inhibitor ibrutinib, in clinical use for the therapy of WM, was added prior to the stimulation with EVs. Decrease of the NF-κB activation (Figure 2F) was detected, corroborating signaling in the cytosol through pathway inhibited by ibrutinib.

EV-transferred MyD88^{L265P} recruits the endogenous MyD88^{wt} to potentiate the signaling

In the previous study we showed that MyD88^{L265P} can recruit the MyD88^{wt}.⁶ This suggested that the introduction of the constitutively active MyD88 mutant into the recipient cells might trigger the inflammatory response by the recruitment of the endogenous MyD88^{wt}. MyD88^{L265P}-CFP containing EVs were therefore prepared in order to monitor trafficking and interactions of the MyD88 in the recipient cells. Incubation of EV_{L265P-CFP} with target HEK293 cells and subsequent addition of membrane dye CT-B showed mainly cytosolic localization of the MyD88 within the recipient cells (Figure 3A). When EV_{L265P-CFP} were added to the HEK293 cells expressing MyD88^{wt-YFP} (Figure 3B), we observed that MyD88^{L265P} and MyD88^{wt} formed mixed aggregates inside recipient cells, demonstrating that the EV-mediated uptake of MyD88^{L265P} enabled its release into the cell cytoplasm, where it interacted with the endogenous protein. Furthermore, upon addition of a mixture of EVs isolated from cells producing MyD88^{L265P-CFP} or MyD88^{L265P-YFP} to the recipient cells, mixed aggregates were observed inside recipient cells (Figure 3C), demonstrating cytoplasmic interaction between proteins that enter the cells via EVs. Similar experiments were performed in MyD88^{KO} cells. MyD88^{KO} HEK293 cell line was prepared by using CRISPR/Cas9 technology (to be published elsewhere and supplemental Figure 3C-D). In wt cells the overexpressed MyD88 is observed in the cytoplasm

in the form of aggregates even at very low expression levels due to the formation of complex with endogenous MyD88, but in MyD88^{KO} cells disperse cytosolic localization of MyD88^{wt-YFP} was observed (white arrows) (Figure 3D). Internalization of EV_{L265P-CFP} led to the formation of aggregates with MyD88^{wt-YFP}. In agreement with this, MyD88^{KO} cell line responded less efficiently to the EV_{L265P} stimulation (Figure 3E, left) than MyD88^{wt} cells (Figure 3E, right), demonstrating that the transmitted MyD88^{L265P} is able to form active myddosomes by recruitment of the endogenous MyD88^{wt}, which potentiates the activation of the signaling pathway.

Macrophages and mast cells respond to the MyD88^{L265P} containing EVs

Efficiency of the EV internalization can strongly differ between cells. EV_{MWCL1} were stained by a lipophilic dye PKH67 and added to several cell types. After 6 h HEK293, THP1, macrophage cell line, bone marrow derived macrophages (iBMDMs) as well as bone marrow derived mast cells (BMCMCs) were able to uptake the EVs, but no uptake was observed by B cells and peripheral blood mononuclear cells (PBMCs) – a minor uptake was probably due to macrophages (Figure 4A and supplemental Figure 4). Interestingly, fusion of the EV_{MWCL} with the plasma membrane was observed in HEK293 cells and BMCMCs (Figure 4D and supplemental Figure 4), which could contribute to the efficient delivery of the myddosomal components into the cytosol, bypassing the endocytosis and phagocytosis. In accordance with this observation no stimulation of B cells with EV_{L265P} was observed (data not shown), but cytokine expression was detected in iBMDMs (Figure 4B-C) as well as in BMCMCs (Figure 4E-F) after treatment with EV_{L265P}, EV_{MWCL} or EV_{BCWM}, implying those cells as the likely targets of the EV delivery, which is particularly relevant for the bone marrow environment.

EVs carrying myddosomal complex modulate bone marrow microenvironment

Clinical manifestations of WM are mainly due to the monoclonal IgM protein and infiltration, predominantly by the lymphoplasmacytic cells into the bone marrow. Mast cell hyperplasia is common and can stimulate WM cell proliferation.³⁴ In WM, blood smears also show a characteristic striking rouleaux formation (reviewed in³⁵). To demonstrate that EVs can remain in the circulation and be internalized by the recipient cells *in vivo* EVs were labelled with PKH67 and injected i.p. or i.v.. After 4 h uptake of EVs was detected in cells from peritoneal lavage (supplemental Figure 5A) as well as in organs such as liver, spleen and lungs (supplemental Figure 5B). Moreover, to show that signaling competent MyD88 can be transmitted into the recipient cells *in vivo*, HEK293 cells expressing NF- κ B dependent luciferase reporter were introduced s.c. into the BALB/c mice. EV_{CONT} or EV_{L265P} were subsequently administered i.v. (150 μ g). After 24 h we were able to detect the luciferase activity in animals only when EV_{L265P} were injected (Figure 5A), confirming that EVs can transmit the MyD88 signal systemically.

As the main pathology of the WM takes place in the bone marrow, PKH67-labelled EVs were injected into the C57BL/6 mice femurs. The uptake by cells in the bone marrow was confirmed by flow cytometer after 16 h (Figure 5B, black line). EV_{MWCL} or EV_{CONT} were injected (150 μ g/mouse) into the femurs to see if the uptake of EVs can change the microenvironment and contribute to the bone marrow pathology. After 8 days all mice had normal levels of total proteins (TP) and albumins (ALB) in the serum (Figure 5C). However, mice injected with EV_{MWCL} but not EV_{CONT} and PBS exhibited increased globulin concentrations (GLOB) (Figure 5C-D) in all mice (5/5), and induced rouleaux formation in blood (red arrow) (Figure 5E) in 2/5 mice. Bone marrow slides were stained with antiCD40L (antiCD154) antibodies for mast cell detection. Increased number of mast cells was detected in bone marrow only in mice that formed the rouleaux (Figure 5F) in comparison to the EV_{CONT} injection, demonstrating that EVs can directly influence the microenvironment.

Finally, the presence of MyD88 in the EVs from samples of patients with WM was investigated to establish the relevance of our observations for pathology. Using differential ultracentrifugation exosomes and MVs were isolated from the bone marrow aspirates of WM patients. MyD88 (Figure 5G) as well as IRAK4 (Figure 5H) were detected in the EVs (MV and exosomes), but not in the serum, confirming the presence MyD88^{L265P} in the EVs under the pathological conditions.

Discussion

Pattern recognition receptors (PRRs) including TLRs and TLR signaling pathways have been implicated in a number of hematological malignancies (reviewed in³⁶) and in other tumors.³⁷⁻³⁹ Although the mechanisms by which inflammation promotes neoplastic transformation are not fully understood, cancer has strong connection to chronic inflammation.^{40,41} Persistent inflammation establishes a microenvironment, which contains macrophages, dendritic cells, natural killer cells and T and B lymphocytes in addition to the surrounding stroma.^{42,43} These diverse cells communicate with each other by means of a direct contact or soluble mediators, which support cell proliferation, survival and resistance to apoptosis thus influencing the tumor formation and growth.^{43,44} EVs are able to mediate the transfer of different kinds of molecules. Due to the small amount of the cellular content transmitted by the EVs, a potent physiological effect can only be expected from the components whose activity is amplified in target cells, such as RNAs, enzymes, transcription factors (TFs) or components that could engage endogenous molecules in the target cells thus triggering the signaling cascade. As an example, TFs NF- κ B and COUP-TF I were detected in MVs released from platelets and transmitted to neutrophils, but activation of target genes in recipient cells has not been investigated.⁴⁵ Here we report the transfer and activation of target cell signaling by the constitutively active MyD88^{L265P} as well as IRAK4 as the components of the myddosomal

complex. We demonstrated that MyD88^{L265P} can be delivered by the EVs and can reach the cytosol using several internalization pathways. Fusion of EVs with plasma membrane enables direct release of the content into the cytoplasm, while upon EV phagocytosis or endocytosis the EVs need to fuse with organelle membranes by a process referred to as back-fusion (reviewed in⁴⁶). We demonstrated that the EV-delivered MyD88^{L265P} engages the endogenous MyD88^{wt} resulting in potentiated activation of the inflammatory pathway, including activation of the NF- κ B and inflammatory cytokine/chemokine production in macrophages and mast cells, both important for the pathogenesis of WM (Figure 6). The presence of the MyD88^{L265P} was detected in bone marrow aspirates of patient samples, where the local concentration of EVs is very high and persistent and which could strongly contribute to the inflammation in the cancer cell microenvironment. Bacterial or viral infection, characterized by activation of TLRs, might also promote the release of MyD88^{wt} in the EVs. However, in proteomic analysis of exosomes released after LPS stimulation of BMDM,⁴⁷ MyD88 has not been detected, suggesting that the dissemination by EVs may be specific to MyD88^{L265P} in certain types of the lymphoma.

By injecting EVs into the bone marrow of mice we detected a significant increase in globulins in all mice, with increased mast cells observed in the bone marrow along with blood rouleaux formation. It has been recently reported⁴⁸, that mice with MyD88^{L252P} (an analog of human L265P) spontaneously developed signs of DLBCL-like neoplasms, but no changes typical for WM such as infiltration of lymphoid cells to the bone marrow were observed. Also no engraftment in the bone was observed when BCWM.1 cells were injected s.c. to immunocompromised mice. Interestingly, the pathology developed when human bone chip from a fetal femur was transplanted into mice, showing that human bone marrow environment was important for the engraftment of tumor cells²⁷ where the EVs might play an important role. It is likely that other constitutively active signaling pathway components that are able to engage endogenous mediators might be transferred between cells and activate signaling in the

recipient cells. ASC adaptor plays a similar role in the inflammasome activation as the MyD88. ASC overexpression leads to the constitutive activation of caspase-1 and formation of large aggregates, called specks, resembling the myddosomal complex. ASC specks have been observed in the extracellular milieu, where they can be taken up by other cells. Those aggregates enabled nucleation of the soluble ASC in the recipient cells and triggered signaling.^{49,50} Although it has been proposed that ASC particles have to be released by cells into the extracellular medium and taken up by target cells by phagocytosis, recently, NLRP3 and ASC were found in the exosomes secreted from the LPS/nigericin stimulated BMDMs.⁴⁷ Interestingly, those exosomes were able to activate NF- κ B signaling and induce pyroptosis, suggesting that the EV-mediated transfer may play an important role as well.

In conclusion, we report that the transfer of constitutively active signaling mediators via EVs represents a new mechanism of intercellular transfer that is amplified in the recipient cells by the recruitment of the endogenous signaling mediators. This process may play an important role in cancer development by triggering inflammation in the nontransformed cells independently of the membrane receptors.

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Authorship

Contribution: MMK designed and performed most of the experiments, DL designed and performed animal experiments, MB and MMK performed confocal microscopy experiments, RR designed and performed electron microscopy, JGC and ZRH isolated EVs from human samples and performed WB. MMK, RJ and SPT wrote the paper.

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References

1. Motshwene PG, Moncrieffe MC, Grossmann JG, et al. An Oligomeric Signaling Platform Formed by the Toll-like Receptor Signal Transducers MyD88 and IRAK-4. *Journal of Biological Chemistry*. 2009;284(37):25404-25411.
2. Gay NJ, Symmons MF, Gangloff M, Bryant CE. Assembly and localization of Toll-like receptor signalling complexes. *Nature Reviews Immunology*. 2014;14(8):546-558.
3. Kawai T, Akira S. TLR signaling. *Cell Death and Differentiation*. 2006;13(5):816-825.
4. von Bernuth H, Picard C, Jin ZB, et al. Pyogenic bacterial infections in humans with MyD88 deficiency. *Science*. 2008;321(5889):691-696.
5. Ngo VN, Young RM, Schmitz R, et al. Oncogenically active MYD88 mutations in human lymphoma. *Nature*. 2011;470(7332):115-U133.
6. Avbelj M, Wolz OO, Fekonja O, et al. Activation of lymphoma-associated MyD88 mutations via allosterically-induced TIR-domain oligomerization. *Blood*. 2014;124(26):3896-3904.
7. Wang JQ, Jeelall YS, Beutler B, Horikawa K, Goodnow CC. Consequences of the recurrent MYD88(L265P) somatic mutation for B cell tolerance. *Journal of Experimental Medicine*. 2014;211(3):413-426.
8. Puente XS, Pinyol M, Quesada V, et al. Whole-genome sequencing identifies recurrent mutations in chronic lymphocytic leukaemia. *Nature*. 2011;475(7354):101-105.
9. Treon SP, Xu L, Yang G, et al. MYD88 L265P Somatic Mutation in Waldenstrom's Macroglobulinemia. *New England Journal of Medicine*. 2012;367(9):826-833.
10. Xu L, Hunter ZR, Yang G, et al. MYD88 L265P in Waldenstrom macroglobulinemia, immunoglobulin M monoclonal gammopathy, and other B-cell lymphoproliferative disorders using conventional and quantitative allele-specific polymerase chain reaction. *Blood*. 2013;121(11):2051-2058.
11. Nagao T, Oshikawa G, Ishida S, et al. A novel MYD88 mutation, L265RPP, in Waldenstrom macroglobulinemia activates the NF-kappa B pathway to upregulate Bcl-xL expression and enhances cell survival. *Blood Cancer Journal*. 2015;5.
12. Yang G, Zhou YS, Liu X, et al. A mutation in MYD88 (L265P) supports the survival of lymphoplasmacytic cells by activation of Bruton tyrosine kinase in Waldenstrom macroglobulinemia. *Blood*. 2013;122(7):1222-1232.
13. Skog J, Wurdinger T, van Rijn S, et al. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nature Cell Biology*. 2008;10(12):1470-U1209.

14. Ratajczak J, Wysoczynski M, Hayek F, Janowska-Wieczorek A, Ratajczak MZ. Membrane-derived microvesicles: important and underappreciated mediators of cell-to-cell communication. *Leukemia*. 2006;20(9):1487-1495.
15. Hugel B, Carmen M, Martinez MC, Kunzelmann C, Freyssinet JM. Membrane microparticles: Two sides of the coin. *Physiology*. 2005;20:22-27.
16. Lopez JA, Kearon C, Lee AY. Deep venous thrombosis. *Hematology Am Soc Hematol Educ Program*. 2004:439-456.
17. Sung BH, Ketova T, Hoshino D, Zijlstra A, Weaver AM. Directional cell movement through tissues is controlled by exosome secretion. *Nature Communications*. 2015;6.
18. Yue SJ, Mu W, Erb U, Zoller M. The tetraspanins CD151 and Tspan8 are essential exosome components for the crosstalk between cancer initiating cells and their surrounding. *Oncotarget*. 2015;6(4):2366-2384.
19. Clancy JW, Sedgwick A, Rosse C, et al. Regulated delivery of molecular cargo to invasive tumour-derived microvesicles. *Nature Communications*. 2015;6.
20. Peinado H, Aleckovic M, Lavotshkin S, et al. Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. *Nature Medicine*. 2012;18(6):883-+.
21. Costa-Silva B, Aiello NM, Ocean AJ, et al. Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver. *Nature Cell Biology*. 2015;17(6):816-+.
22. Fevrier B, Vilette D, Archer F, et al. Cells release prions in association with exosomes. *Proceedings of the National Academy of Sciences of the United States of America*. 2004;101(26):9683-9688.
23. Skogberg G, Gudmundsdottir J, van der Post S, et al. Characterization of Human Thymic Exosomes. *Plos One*. 2013;8(7).
24. Ho AW, Hatjiharissi E, Ciccarelli BT, et al. CD27-CD70 interactions in the pathogenesis of Waldenstrom macroglobulinemia. *Blood*. 2008;112(12):4683-4689.
25. Elswa SF, Novak AJ, Ziesmer SC, et al. Comprehensive analysis of tumor microenvironment cytokines in Waldenstrom macroglobulinemia identifies CCL5 as a novel modulator of IL-6 activity. *Blood*. 2011;118(20):5540-5549.
26. Hodge LS, Novak AJ, Grote DM, et al. Establishment and characterization of a novel Waldenstrom macroglobulinemia cell line, MWCL-1. *Blood*. 2011;117(19):E190-E197.
27. Santos DD, Ho AW, Tournilhac O, et al. Establishment of BCWM.1 cell line for Waldenstrom's macroglobulinemia with productive in vivo engraftment in SCID-hu mice. *Experimental Hematology*. 2007;35(9):1366-1375.
28. Zilber S, Epstein N, Lee SW, et al. Mouse femoral intramedullary injection model: Technique and microCT scan validation. *Journal of Biomedical Materials Research Part B-Applied Biomaterials*. 2008;84B(1):286-290.
29. Perez JM, Chirieleison SM, Abbott DW. An IkappaB Kinase-Regulated Feedforward Circuit Prolongs Inflammation. *Cell Rep*. 2015;12(4):537-544.
30. Parolini I, Federici C, Raggi C, et al. Microenvironmental pH Is a Key Factor for Exosome Traffic in Tumor Cells. *Journal of Biological Chemistry*. 2009;284(49):34211-34222.
31. Feng D, Zhao WL, Ye YY, et al. Cellular Internalization of Exosomes Occurs Through Phagocytosis. *Traffic*. 2010;11(5):675-687.
32. Tian T, Zhu YL, Zhou YY, et al. Exosome uptake through clathrin-mediated endocytosis and macropinocytosis and mediating miR-21 delivery. *J Biol Chem*. 2014;289(32):22258-22267.
33. Jefferies CA, Doyle S, Brunner C, et al. Bruton's tyrosine kinase is a Toll/interleukin-1 receptor domain-binding protein that participates in nuclear factor kappaB activation by Toll-like receptor 4. *J Biol Chem*. 2003;278(28):26258-26264.

34. Tournilhac O, Santos DD, Xu L, et al. Mast cells in Waldenstrom's macroglobulinemia support lymphoplasmacytic cell growth through CD154/CD40 signaling. *Annals of Oncology*. 2006;17(8):1275-1282.
35. Stone MJ, Pascual V. Pathophysiology of Waldenstrom's macroglobulinemia. *Haematologica-the Hematology Journal*. 2010;95(3):359-364.
36. Wang JQ, Jeelall YS, Ferguson LL, Horikawa K. Toll-like receptors and cancer: MYD88 mutation and inflammation. *Frontiers in Immunology*. 2014;5.
37. Maeda S, Kamata H, Luo JL, Leffert H, Karin M. IKK beta couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. *Cell*. 2005;121(7):977-990.
38. Swann JB, Vesely MD, Silva A, et al. Demonstration of inflammation-induced cancer and cancer immunoediting during primary tumorigenesis. *Proceedings of the National Academy of Sciences of the United States of America*. 2008;105(2):652-656.
39. Oblak A, Jerala R. Toll-like receptor 4 activation in cancer progression and therapy. *Clin Dev Immunol*. 2011;2011:609579.
40. Grivennikov SI, Greten FR, Karin M. Immunity, Inflammation, and Cancer. *Cell*. 2010;140(6):883-899.
41. Munn LL. Cancer and inflammation. *Wiley Interdiscip Rev Syst Biol Med*. 2016.
42. de Visser KE, Eichten A, Coussens LM. Paradoxical roles of the immune system during cancer development. *Nature Reviews Cancer*. 2006;6(1):24-37.
43. Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. *Cell*. 2011;144(5):646-674.
44. Allavena P, Germano G, Marchesi F, Mantovani A. Chemokines in cancer related inflammation. *Experimental Cell Research*. 2011;317(5):664-673.
45. Duchez AC, Boudreau LH, Naika GS, et al. Platelet microparticles are internalized in neutrophils via the concerted activity of 12-lipoxygenase and secreted phospholipase A2-IIA. *Proc Natl Acad Sci U S A*. 2015;112(27):E3564-3573.
46. Cocucci E, Meldolesi J. Ectosomes and exosomes: shedding the confusion between extracellular vesicles. *Trends Cell Biol*. 2015;25(6):364-372.
47. Zhang Y, Liu F, Yuan Y, et al. Inflammasome-Derived Exosomes Activate NF-kappaB Signaling in Macrophages. *J Proteome Res*. 2017;16(1):170-178.
48. Knittel G, Liedgens P, Korovkina D, et al. B-cell-specific conditional expression of Myd88(p.L252P) leads to the development of diffuse large B-cell lymphoma in mice. *Blood*. 2016;127(22):2732-2741.
49. Baroja-Mazo A, Martin-Sanchez F, Gomez AI, et al. The NLRP3 inflammasome is released as a particulate danger signal that amplifies the inflammatory response. *Nature Immunology*. 2014;15(8):738-+.
50. Franklin BS, Bossaller L, De Nardo D, et al. The adaptor ASC has extracellular and 'prionoid' activities that propagate inflammation. *Nature Immunology*. 2014;15(8):727-+.

Figure Legends

Figure 1. MyD88^{L265P} is secreted from the cells in EVs, which can activate signaling pathway in recipient cells. EV_{L265P} and EV_{CONT} were isolated and (A) MyD88 (protein load 10 µg) and Tsg101 or (B) IRAK4 (protein load 30 µg) and Tsg101 were detected by WB. (C) HEK293 cells expressing luciferase under NF-κB promotor and *Renilla* luciferase for normalization were stimulated with EV_{L265P} and EV_{CONT} (12 µg/ml) for 24 h. EV_{MWCL} were isolated and (D) MyD88 (protein load 10 µg) and Tsg101 or (F) IRAK4 (protein load 30 µg) and Tsg101 were detected by WB. (E) EV_{MWCL} were isolated, fixed and sections were prepared for immune-labeling with antiMyD88 Ab and protein A conjugated to 10 nm gold. TEM was performed. (G) HEK293 cells expressing luciferase under NF-κB promotor and *Renilla* luciferase for normalization were stimulated with EV_{MWCL} and EV_{CONT} (80 µg/ml) for 24 h. Cells were stimulated for 24 h. Negative controls are transfected but unstimulated cells. Dual luciferase tests for NF-κB activity were performed.

Figure 2. Internalization of the intact EVs is necessary for signaling. EV_{L265P} were isolated either from the NEMO^{KO} cells or NEMO^{wt} cells. (A) HEK293 cells expressing luciferase under the NF-κB promotor and *Renilla* luciferase for normalization were stimulated with EV_{L265P} (12 µg/ml) for 24 h. EV_{L265P} were isolated from HEK293 cells and (B) IRAK4^{wt} and IRAK4^{KO} BMDMs were stimulated for 16 h (30 µg/ml). LPS was a positive control (10 ng/ml). RNA was isolated and qPCRs for *Rantes* was performed. (C) EV_{L265P} and EV_{CONT} were submitted to 3 frozen/thaw cycles and then added to cells (12 µg/ml). (D-F) Dynasore (30 and 50 µM), LY294002 (50 µM) or ibrutinib (4 µM) were added to cells 1 h prior stimulation with EV_{L265P} and EV_{CONT} (12 µg/ml) or EV_{MWCL} (80 µg/ml) and EV_{CONT} (80 µg/ml). HEK293 cells expressing luciferase under NF-κB promotor and *Renilla* luciferase for normalization

were stimulated for 24 h. Negative controls are transfected but unstimulated cells. Dual luciferase tests for NF- κ B activity were performed.

Figure 3. MyD88^{L265P} transferred by the EVs forms aggregates with endogenous MyD88 in the recipient cells. (A) EV_{L265P-CFP} were isolated, added (5 μ g/ml) to the HEK293 cells for 16 h. Cell membranes were dyed with CT-B Alexa647. (B,D) HEK293 and HEK293 MyD88^{KO} cells were transfected with MyD88^{wt-YFP} or (C) left untreated. After 4 h EV_{L265P-CFP} or EV_{L265P-YFP} (both 5 μ g/ml) were added for 16 h. Confocal imaging was performed. Bar 10 μ m. (D) HEK293 and HEK293 MyD88^{KO} cells expressing luciferase under NF- κ B promotor and *Renilla* luciferase for normalization were stimulated with EV_{L265P} and EV_{CONT} (12 μ g/ml) for 24 h. Negative controls are transfected but unstimulated cells. Dual luciferase test for NF- κ B activity was performed.

Figure 4. Macrophages and mast cells respond to EV treatment. PKH67 labelled EV_{MWCL} were added to iBMDMs (A) and BMCMCs (D) to observe EV internalization. iBMDMs (B,C) were seeded and stimulated with EV_{CONT}, EV_{L265P} (30 μ g/ml) or EV_{MWCL}, EV_{BCWM} and EV_{Ramos} as negative control (100 μ g/ml) and LPS as positive control (10 ng/ml) for 16 h. BMCMCs (E,F) were seeded and stimulated with EV_{CONT}, EV_{L265P} (30 μ g/ml) or EV_{MWCL} and EV_{Ramos} as negative control (100 μ g/ml) and LPS as positive control (50 ng/ml) for 16 h. RNA was isolated and qPCRs for *Rantes* and *Il6* were performed.

Figure 5. EVs are internalized *in vivo* and modify the bone marrow microenvironment. (A) HEK293 cells were transfected with luciferase under NF- κ B promotor for 24 h. 2 x 10⁶ cells/mouse were injected s.c. After 30 min EV_{CONT} or EV_{L265P} (150 μ g/mouse) were injected i.v. and incubated for 24 h. Luciferin was added and after 10-15 min luminiscence was

measured. PKH67 labelled EVs (100 μ g) were injected into bone marrow. After 16 h uptake of EVs by bone marrow cells was measured (B). EV_{CONT} or EV_{MWCL} (150 μ g) were injected into bone marrow. After 8 days blood was taken to measure albumin, total protein and globulin (C and D) concentrations and blood smears (E) for detection of rouleaux formation were prepared. (F) Tissue sections from femurs were prepared and stained for the presence of mast cells (CD40L). MVs and exosomes were isolated from bone marrow aspirates of untreated WM patients. MVs, exosomes, serum and BCWM.1 cell lysate as positive control were used for MyD88 (G) and IRAK4 (H) detection by WB. GAPDH was used as a positive control for EVs and cell lysate.

Figure 6. Schematic representation of EV-mediated MyD88 signaling. Lymphoma cells release IgMs and cytokines as well as myddosome containing exosomes and MVs. Recipient cells such as macrophages and mast cells uptake EVs and myddosomes are released into the cytosol, where MyD88^{wt} can be recruited. Myddosomes induce inflammation without receptor activation thus changing tumor`s microenvironment.

Figure 1

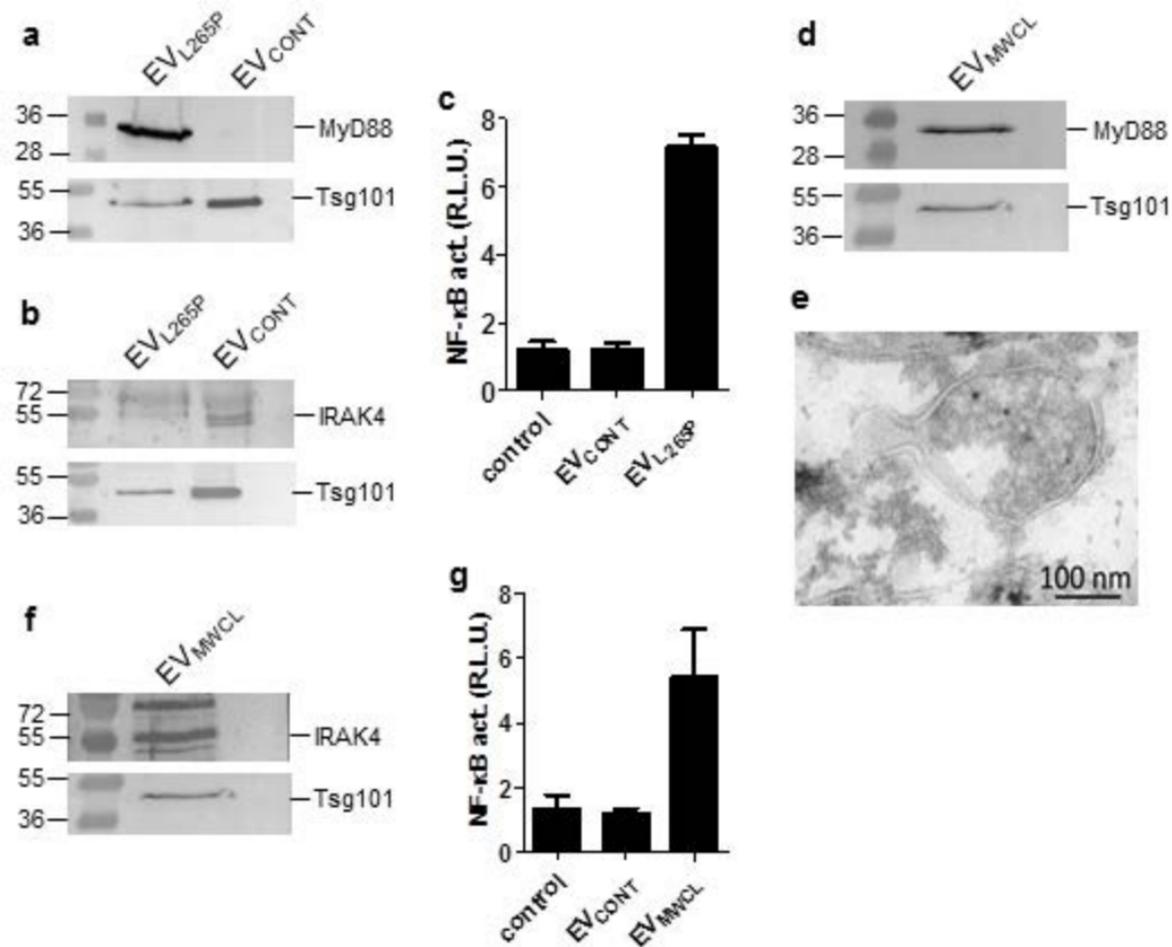


Figure 2

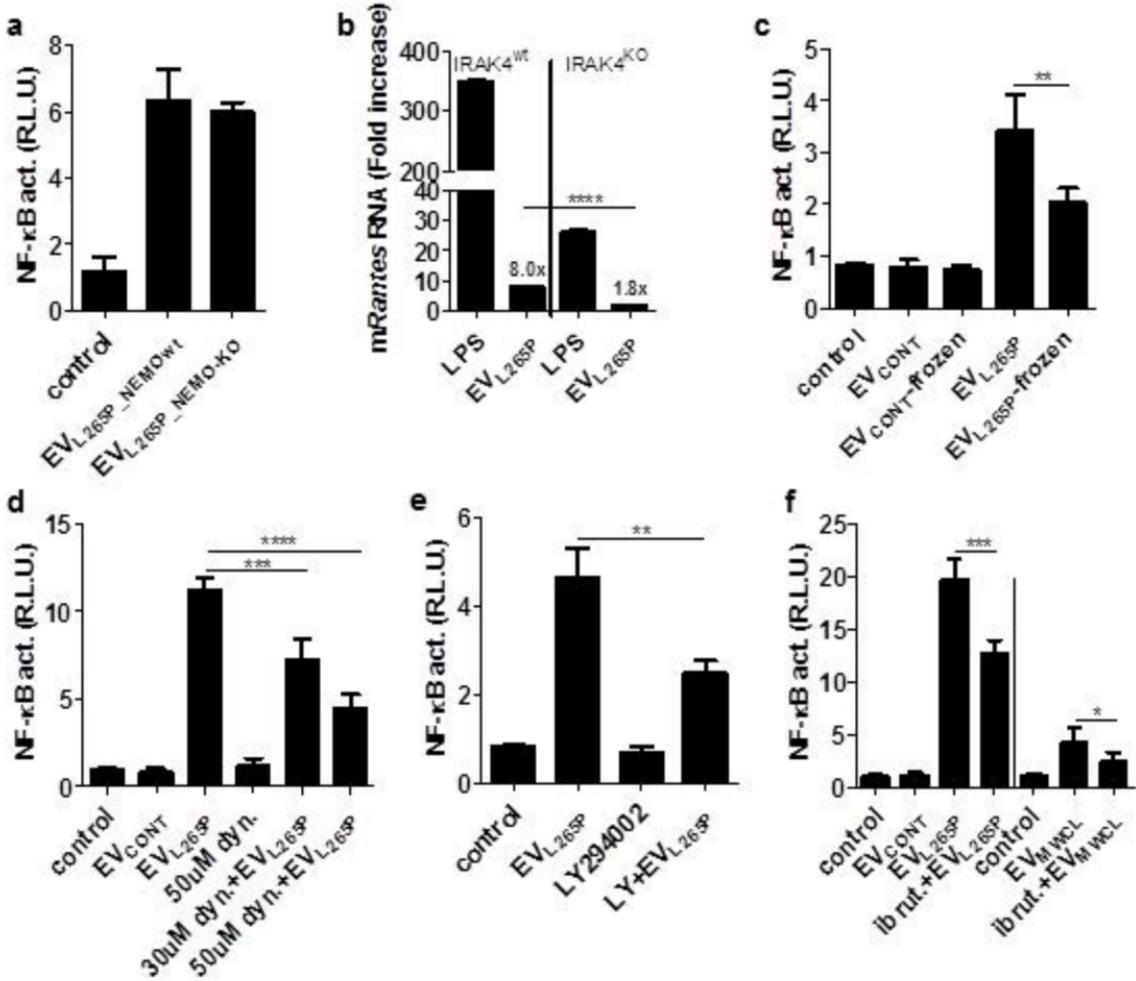
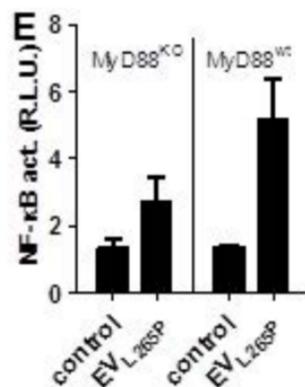
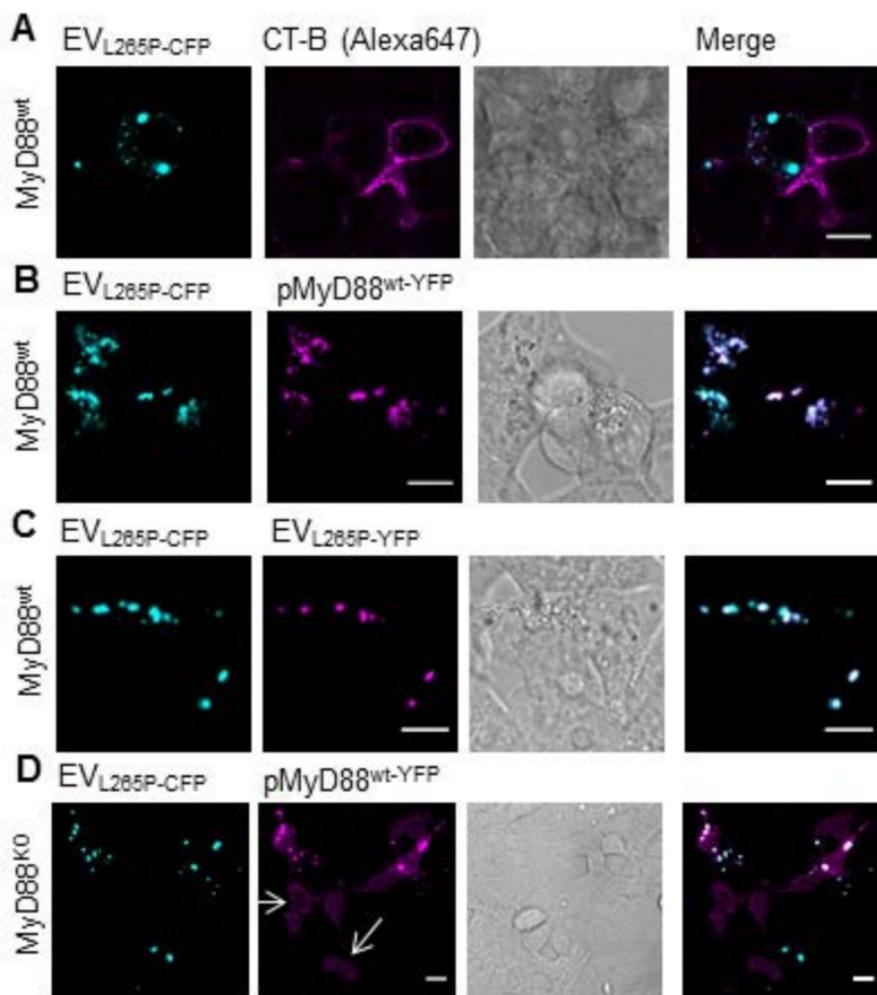
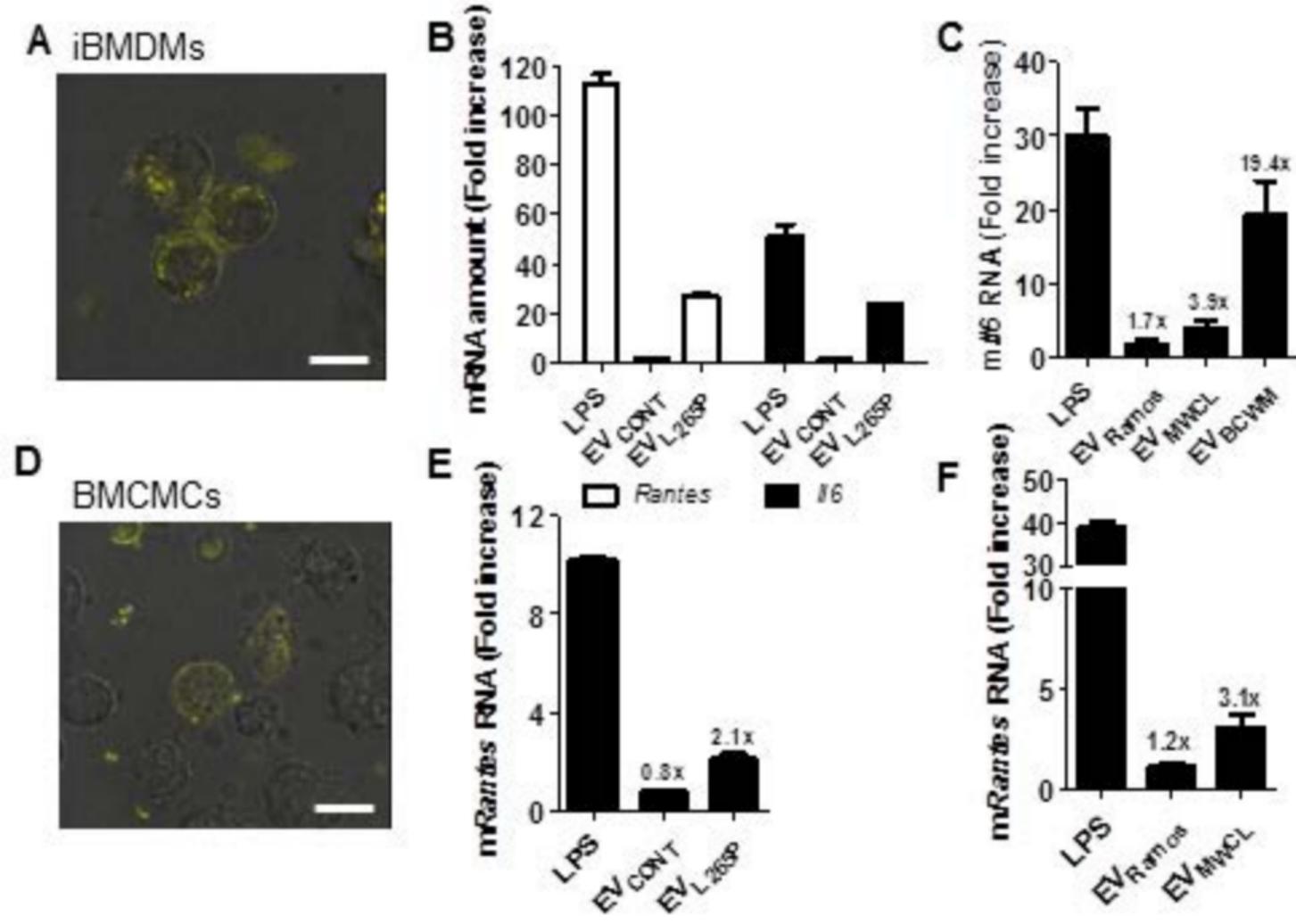


Figure 3





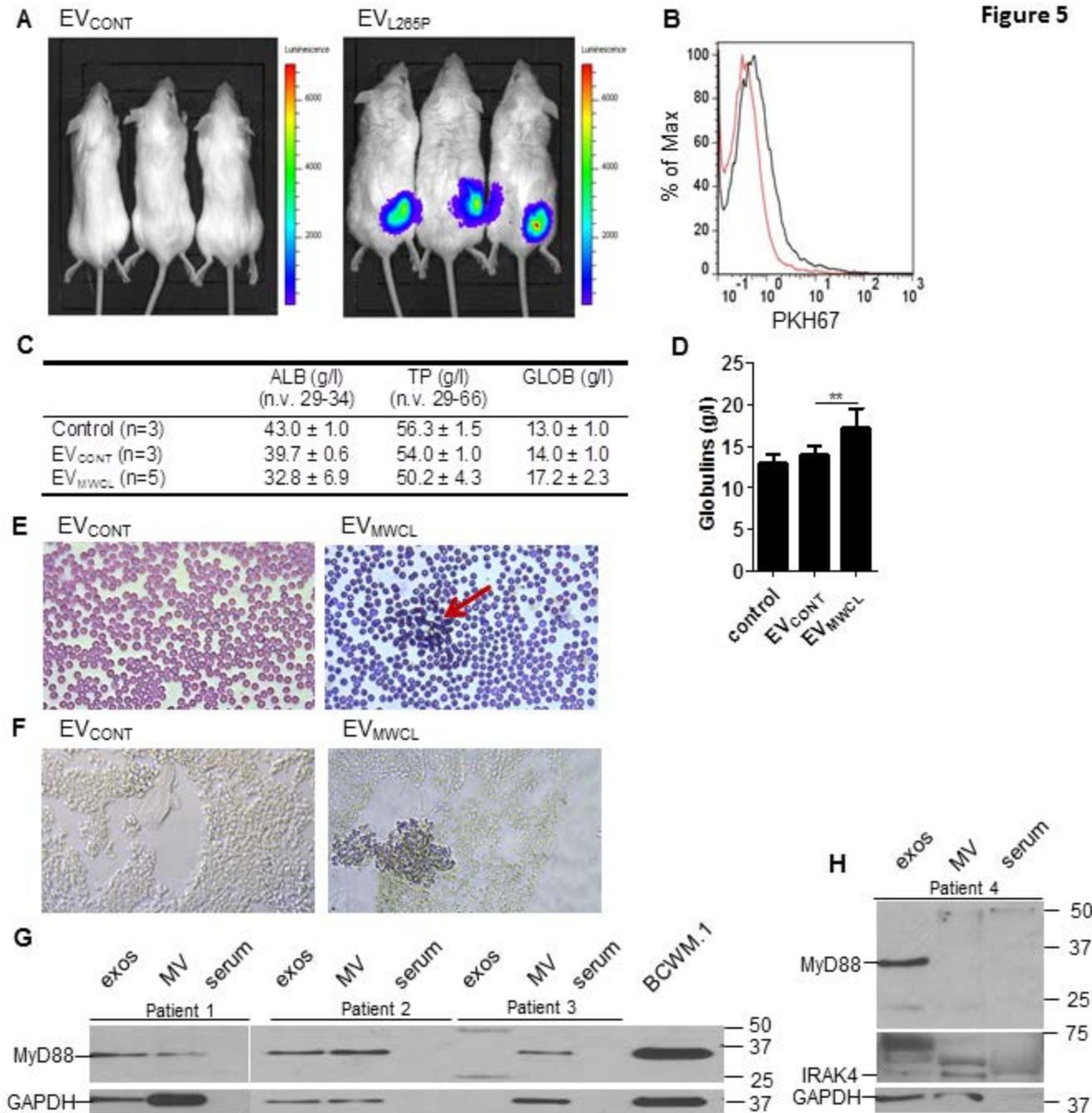
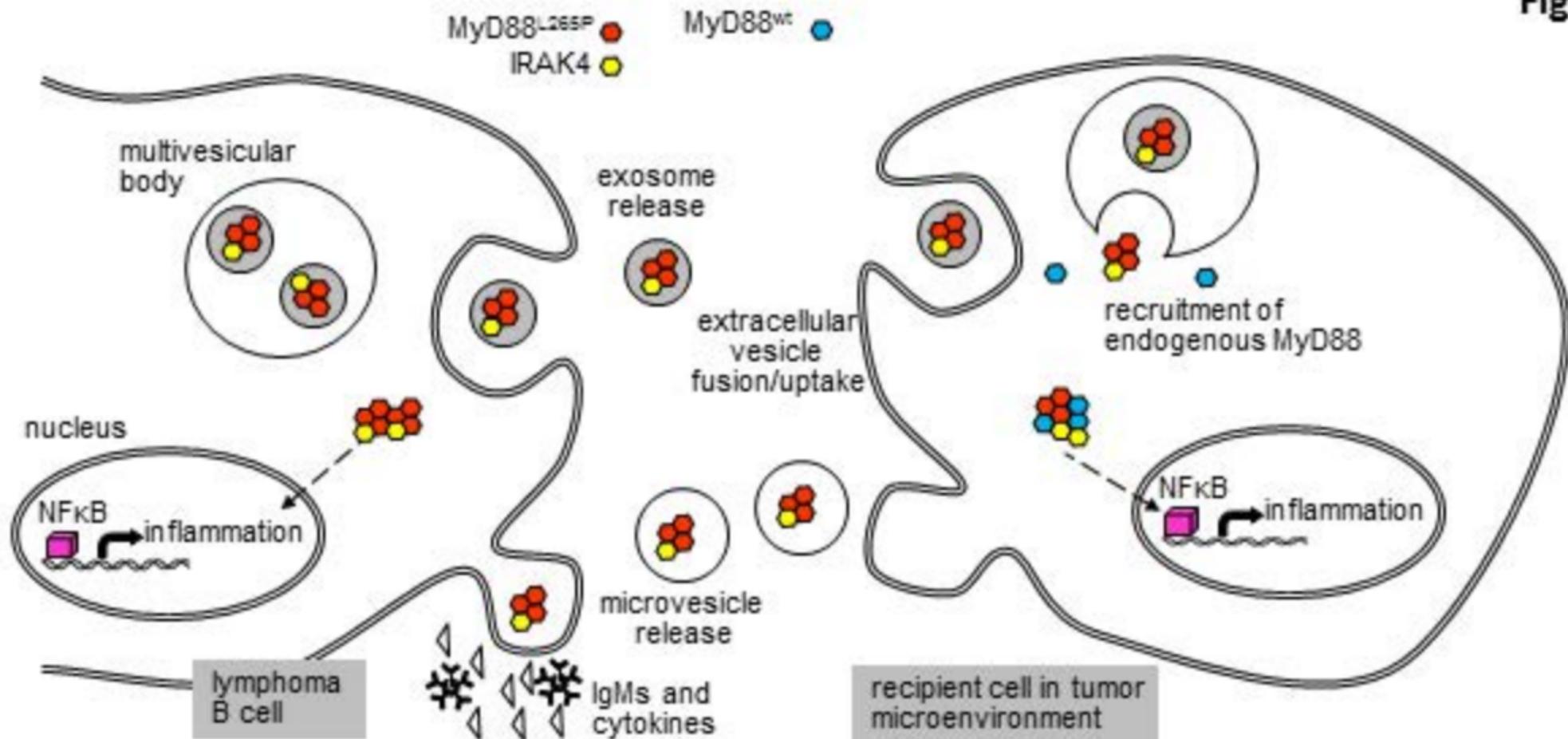


Figure 6





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Extracellular vesicle-mediated transfer of constitutively active MyD88^{L265P} engages MyD88^{wt} and activates signaling

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